RESEARCH ARTICLE



From isolation to connectivity: the effect of floodplain lake restoration on sediments as habitats for macroinvertebrate communities

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Abstract

The present study aimed to identify potential macrozoobenthic habitat indicators of the ecological success of restoration projects. As a part of the complex restoration project in the Słupia River floodplain (N Poland), the connectivity between three oxbow lakes and the river channel was re-established to improve biodiversity of the floodplain area, including bottom fauna. Following restoration, changes in the dynamics of flowing water and water levels induced the transformation of oxbows from plesiopotamal (lentic) to eupotamal (lotic) and subsequently to parapotamal (semi-lotic) habitats. The restored sites underwent a rapid depletion of benthic coarse particulate organic matter, with direct changes in most of the investigated parameters of bottom sediments, including conductivity, total organic carbon, soluble reactive phosphorus and total nitrogen. Redundancy analysis revealed that the changes in benthic fauna structure resulted from the increased connectivity with the river (flow rate) and changes in the chemical properties of sediments. The restored oxbow lakes were colonized by six new macroinvertebrate species whose density increased substantially. The assessment of the overall river-floodplain system restoration project indicated positive implications for improving the qualitative and quantitative structure of benthic fauna. However, to confirm ecologically successful restoration, it is necessary to evaluate appropriate sets of indicators based on a complex food web structure and more efficient or enhanced ecosystem functions. This study contributes to the discussion of sustainable management of floodplains to provide benefits to macroinvertebrates as indicators of aquatic ecosystem health under different restoration activities.

Keywords Benthic invertebrates · Floodplain · Bottom sediments · Restoration

Introduction

The habitat heterogeneity of aquatic ecosystems has been recognized as an important factor promoting biological diversity, particularly that of invertebrate (Schiemer et al.

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1999; Buijse et al. 2005) and fish communities (Miranda et al. 2014; Dolédec et al. 2015). Areas with the highest biodiversity potential include floodplain river ecosystems. Under natural conditions, floodplains comprise a mosaic of habitats subjected to a continuum of hydrological conditions in which side arms, oxbow lakes, and paleomeanders, which represent habitats of various degrees of connectivity, are arrayed along both sides of the river bed, providing 'hot-spots' of biodiversity (Ward et al. 2002). These habitats serve as flow channels, particularly during periods of high water, and offer reproduction sites for aquatic, amphibian and terrestrial organisms (Reckendorfer 2004).

The tendency of meandering rivers to create bends due to erosion and accumulation processes is part of the natural transformation of river channels. However, accumulating evidence indicates that many floodplains in Europe and North America have been disrupted due to straightening and embankment of river channels. Past practices have resulted in the disconnection of floodplains from rivers, with a loss in biodiversity and a demand for complex restoration work. These hydrotechnical projects have led to permanent isolation of former meander bends, consequently disrupting natural processes of water exchange between cut-offs and the river channel. High and rare flood events appear to be crucial to generate floodplain habitat diversity (Amoros and Bornette 2002) and to allow organisms to exploit stagnant waters as a result of adaptive strategies. A key factor linking river-floodplain system (RFS) properties and the structure of floodplain biota is hydrologic variability, as described by the "flood pulse concept" (Junk et al. 1989). The degree of connectivity of floodplain water bodies is mirrored by the substrate conditions: highly organic mud is typical of stagnant isolated water bodies, whereas lotic water bodies feature coarse, highly permeable mineral deposits.

Regardless of the cause, a lack of habitat regeneration by fluvial dynamics leads to the disappearance of lentic ecosystems. Cut-offs quickly become filled with sediments as a result of autogenic processes and/or allogenic processes, i.e., accumulation of the material carried by the river during inundation events and by ground water (Sieczko and Peduzzi 2014). Soon after, the isolated water bodies undergo eutrophication and then terrestrialization (Obolewski et al. 2016; Palmer et al. 2005). The lakes accumulate organic matter and become shallower, with decreasing biodiversity due to progressive domination of species tolerant of the changes in environmental quality (e.g., Obolewski 2011; Paillex et al. 2015).

The properties of the bottom substratum determine the abundance and diversity of benthic macroinvertebrates (Culp et al. 1983; Rempel et al. 2000; Obolewski and Glińska-Lewczuk 2013). However, macrozoobenthos influence bottom sediments (Jones and Jago 1992; Żbikowski and Kobak 2007) and improve water exchange by increasing oxygen levels above the sediment (Svensson and Leonardson 1996). The rate and direction of matter exchange between water and bottom sediments are also influenced by the abundance and taxonomic composition of zoobenthos (Covich et al. 1999). Restoration tasks within a single RFS that rely mainly on the reconnection of aquatic ecosystems with the river channel influence habitat diversity measured on the scale of a single sampling site (α —diversity), ecosystem (β —diversity) or floodplain (γ —diversity).

Floodplain lake restoration refers mainly to changes in the pathways of water, resources and organisms between water bodies. How the interaction between water flow intensity and direction influences the turnover of macroinvertebrates in floodplain systems remains unclear, as does the magnitude, pathway, and distance through which the floodwater transports the biota, particularly macroinvertebrates, to newly inundated habitats in a lateral connectivity gradient (Mesa et al. 2012).

Our previous studies (Obolewski and Glińska-Lewczuk 2013; Obolewski et al. 2016) demonstrated that the restoration effects in RFSs can be analyzed at different temporal scales. Short-term effects are observed soon after reconnection when the system is not stable and the structures of the sediment, water quality, and biota (here: macrozoobenthos) vary significantly between seasons, whereas long-term effects, usually on a multi-year scale, may be assessed when the biota attains relative multiseasonal stability.

Distinct spatial gradients in sediment structure are destroyed within a single floodplain lake during flushing with river water. Activation of water leads to the resuspension of bottom sediments and the release of accumulated nutrients into the water. This sudden change in habitat conditions is a source of stress for hydrobionts, particularly highly sensitive benthic macroinvertebrates. We have demonstrated (Obolewski and Glińska-Lewczuk 2013; Obolewski et al. 2016) that water flowing into floodplain lakes carries suspended matter that is deposited at the bottom, contributing to lake shallowing and inhibiting connectivity in the RFS.

In the current study, we observed changes in floodplain lakes at 3 and 5 years after restoration, and these observations were used to test the following hypotheses: (1) the chemical composition of bottom sediments in floodplain lakes is modified in response to reconnection with the river channel; (2) the observed changes subside gradually as the flow between the floodplain lake and the river is restricted to create a semi-lotic system; (3) the density and species richness of macroinvertebrates are largely determined by the physicochemical properties of bottom sediments; (4) oneside connectivity of floodplain lakes with the river channel plays a key role in restoration processes by creating a distinct gradient in sediment structure along a floodplain lake to support microhabitat heterogeneity for various groups of macroinvertebrates. Full hydrological connectivity (both arms) of the RFS plays a less important role.

Materials and methods

Study sites

The surveyed area occupies the central part of the Słupia River floodplain, which features numerous water bodies (upstream OLS1: N 54°23'0.0649"/E17003'0.22; down-stream OLS 3: N54°23'0.2811"/E17°02'0.0166"). The river is one of the largest watercourses feeding the southern Baltic Sea (catchment area—1620 km², length—138.6 km, mean annual flow rate—15.5 m³ s⁻¹; flows exceeding 100 m³ s⁻¹ are generally noted between October and March) (Obolewski 2011). The Słupia River floodplain is 0.3–0.4 km wide. Its meandering section spreads out over 19 km between the village of Lubuń and the town of Słupsk (N Poland). The

floodplain area is covered mostly by meadows and pastures, which are used for agricultural purposes, and 15.6% of its area is occupied by wetland ecosystems. In the early twentieth century, the river channel was artificially regulated, and its meanders were cut off, which led to the formation of nearly 50 oxbow lakes in the floodplain on both sides of the main stream along a 20-km section.

In the summer of 2008, water flow was restored in three floodplain lakes by dredging, and two of those water bodies were merged (Fig. 1). Two cut-off (oxbow) lakes (OLS1 and OLS2) and one parapotamal lake (OLS3) were selected for the restoration project. All three water bodies were smaller than 1 hectare, and their maximum depth did not exceed 1.5 m. Between 2008 and 2011, the analyzed lakes constituted lotic water bodies, but in 2012, the upstream arm began to fill with suspended matter, and the lakes were gradually transformed into semi-lotic water bodies (Obolewski et al. 2016).

The first series of samples was collected before hydrotechnical work started in the summer of 2007. After the disruptive effects of the restoration measures had subsided, the second series of samples was collected in the summer of 2011. The third batch was sampled in the summer of 2013 soon after water flow in the upstream arm of the river became blocked. The above approach supported a comparison of changes resulting from the loss of connectivity in the RFS. Samples of benthic sediments and benthic macroinvertebrates were collected four times in each year of the study, in April, June, August and October. Sampling took place in the upstream and downstream arms and in the center of floodplain lakes (each N=72) as well as in a reference point in the river (N=12). The merged lakes OLS1 and OLS2 were regarded as a single body of water.

Hydrological analyses

River gauge levels and discharges were measured in Słupsk (33.5 km from the river mouth). Discharge (Q) was calculated as the product of the cross-section of the channel or basin of the oxbow lake and average flow velocity in the cross-sectional area. Flow velocity in oxbows was measured weekly during the study period using an electromagnetic



Fig. 1 Location of study area and changes of restored floodplain lakes in 2007 (1 year before restoration) and in 2011 and 2014 (3 and 6 years after restoration)

flow meter (model 801-flat type; Valeport, UK). Gauge levels were recorded every hour using three automatic data loggers (Mini-Diver, Van Essen Instruments, Holland) positioned in each analyzed water body. Gauge levels were compensated by changes in atmospheric pressure with a Baro-Diver data logger (Van Essen Instruments, Holland).

Physicochemical analyses

Physicochemical variables of sediments were selected for relation with the hydrological connectivity of the floodplain lakes. Nine sediment samples were captured using an Ekman type dredge (Eijkelkamp, Holland) from the upstream, downstream and deepest sites in each of the three restored oxbow lakes. To evaluate the grain size as well as physical and chemical sediment composition, the samples were taken from the surface layer (ca. 0-20 cm).

Bulk density (BD) was calculated as dry mass by volume (g cm^{-3}), and organic matter (OM) was estimated by loss on ignition (LOI) in a muffle furnace (450 °C for 6 h) and expressed as a percentage of sediment dry mass (DM). Particle size distribution was measured aerometrically by Casagrande's method as modified by Prószyński to assess the proportions of clay (<2 μ m), silt (2–63 μ m) and sand $(>63 \mu m)$. Reaction pH was measured potentiometrically both in KCl and H₂O, and conductivity (EC) was measured using an HQ40D (HACH, USA). Calcium carbonate was determined with the gasometric method using Scheibler's device following digestion of sediment in H_2SO_4 and H_2O_2 in an MLS-1200 MEGA microwave digestion unit (method according to HACH, Loveland, Colorado, USA). Phosphorus was photometrically measured as soluble reactive phosphorus (SRP) using the molybdenum blue method. This method employs a single-color reagent consisting of an acidified solution of ammonium molybdate, ascorbic acid and antimony-tartrate. The blue phosphomolybdenum complex formed is read colorimetrically through a 50 mm pathlength flowcell at 880 nm (Murphy and Riley 1962). The specific automated method used is described in Technicon (1973) with the following modifications. Nitrogen (TN) was measured as ammonium NH₄-N after Nessler reaction at 425 nm (HACH). Magnesium (Mg), potassium (K) and sodium (Na) were analyzed using a PerkinElmer (AAnalyst 200) atomic absorption spectrophotometer. Total organic carbon (TOC) was measured with a TOC-analyzer 5000A (Shimadzu, Japan). Each measurement was performed in triplicate.

Fauna collection

Analyses of aquatic invertebrates were limited to benthic invertebrates. Three subsamples were collected with an Ekman grab (sample area: 225 cm^2) at the sampling sites of the bottom sediments. Sampling of benthic fauna was

performed simultaneously with sediment sampling. Samples were rinsed in a sieve with a 0.5-mm mesh size and fixed in 4% formaldehyde solution. In the laboratory, invertebrates were separated from sediments and identified to genus or species level, excluding Oligochaeta (phylum level). The results were expressed in terms of 1 m^2 of lake bottom area. The identified species were classified to the functional feeding group (FFG) according to the classification proposed by Merritt and Cummins (2007) and Willacker et al. (2009):

- Shredders (Sd)—Crustacea (*Gammarus pulex* L., *Asellus aquaticus* L.), Gastropoda (*Viviparus contectus* Millet);
- 2. Gatheringcollectors (GC)—Diptera larvae (*Chironomus* sp., *Sergentia* sp.), Oligochaeta;
- Grazer scrapers (GS)—Ephemeroptera larvae (*Cloëon dipterum* L., *Ephemera danica* Müller), Diptera larvae (*Polypedilum sordens* Wulp.);
- Predators (Pd)—Hirudinea (Erpobdella octoculata L., Glossiphonia complanata L., Helobdella sp.), Megaloptera (Sialis fuliginosa L.), Odonata larvae (Libellula quadrimaculata Feb., Lestes sp.), Coleoptera (Dytiscus marginalis L., Cymbiodyta sp.), Heteroptera (Nepa cinerea L., Corixa sp., Heterocorixa sp., Sigara sp.), Diptera larvae (Procladius sp., Chaoborus sp.);
- Filtering collectors (FC)—Bivalvia (Anodonta anatina L., Pisidium sp., Sphaerium sp.);

The results were used to calculate $D\alpha$ Shannon's diversity index (H'), Pielou's evenness index (J') and D_{β} (Whittaker index) in the Past v. 3.01 program. Gamma-diversity was calculated using the formula: $D\gamma = D\alpha \times D\beta$.

Data analysis

For statistical analysis, floodplain lakes were classified based on the degree of connectivity with the river according to the typology proposed by Amoros and Roux (1988). The final dataset contained basic environmental variables: year of analysis combined with the degree of connectivity phase between an oxbow lake and the river (1—*plesiopotamal phase*, 2—*eupotamal*; 3—*parapotamal*), water flow, physicochemical parameters of bottom sediments, and abundance of benthic macroinvertebrates.

To evaluate general differences in macrozoobenthos structure, we analyzed water bodies (OLS1&2, OLS3) in the studied years and performed analysis of variance (ANOVA) with the Kruskal–Wallis test (K–W) followed by Dunn's test in GraphPad Prism 5.01 (GraphPad Software, USA). The following variables were determined: discharges, physicochemical parameters of bottom sediments, mean density of invertebrates, species richness of invertebrates, D α diversity Shannon's index (H') and Pielou's evenness index (J'). We obtained groupings of species and plots based on benthic fauna community composition using two-way hierarchical agglomerative cluster analyses in PC-ORD 6.08 (McCune and Mefford 2011). We performed two-way cluster analysis independent of group samples and species and then combined these analyses into a single diagram to permit observation of the associations between groups of sample units and species. The data were relativized by species maximum to diminish but not eliminate the influence of species totals on species clustering. We used Ward's method as the linkage method on Euclidean distances.

The dataset was log(x + 1) transformed (ter Braak and Smilauer 2002), and the results were checked for normal distribution (Shapiro-Wilk test) and homoscedasticity (Leven's test). Principal component analysis (PCA) was performed using the covariance method to determine the relative significance of environmental factors in explaining the variability of the tested samples. The redundancy between environmental predictors was determined by redundancy analysis (RDA) considering the variance inflation factor (VIF). Independent variables with VIF > 10 were eliminated from further analyses because they supplied redundant information. The length of the gradient on the F1 axis was determined by detrended correspondence analysis (DCA). The results were non-monotonic when the standard deviation for the analyzed variables was greater than 2 (ter Braak 1995). Consequently, the relationships between environmental parameters (year of analysis, site, discharge, physicochemical properties of sediments) and density of benthic fauna were analyzed by RDA (ter Braak 1995). The statistical significance of canonical axes was determined in the Monte Carlo permutation test (Legendre and Legendre 2012). A subset of independent variables representing the relationships between environmental factors and the species/taxa of benthic fauna was identified by eliminating factors that were not significant for the model. Data were processed statistically in Canoco 4.5 software at probability levels of p < 0.05 and p < 0.01.

The General Linear Model (GLM) was used to detect differences in feeding groups of benthic organism abundances and α -diversity in consecutive periods of lake monitoring and in the selected RDA-significant environmental variables. We used a Poisson error distribution to construct the model and checked for over dispersion of the data (Crawley 2007). The procedure was performed using Past v.3.01c software.

Results

Hydrological conditions

In 2011 and 2014, gauge levels in the Słupia River ranged from 13.20 to 14.21 m above sea level (a.s.l.), and average discharge (Q_{AV}) was estimated as 14 m³ s⁻¹. Inundation did



Fig. 2 a Hydrograph of average daily water levels and flows of the Shupia River at a gauge in Shupsk in the period between April and October 2007–2014. **b** Hydrograph of water flow in the studied flood-plain lakes in the period of study

not occur in the analyzed period. In the Słupia River floodplain, inundation occurs when water levels exceed 14.25 m a.s.l and when the river flow exceeds 22 m³ s⁻¹ (Fig. 2a). The highest water levels in the studied site were noted in July 2007 due to heavy rainfall. The lowest water levels were observed in early fall (October). The floodplain lakes are directly connected to the main river channel; therefore, variations in lake levels and discharges are affected by the river. At the beginning of the study (July 2007), the lakes OLS1 and OLS 2 did not maintain surface connectivity with the river, whereas OLS3 was slightly connected to the river by its downstream arm ($Q_{av} = 0.06 \text{ m}^3 \text{ s}^{-1}$). Consequently, all oxbow lakes were treated as plesiopotamal water bodies. The floodplain lakes became eupotamal habitats in July 2011 due to hydrotechnical work but were transformed into parapotamal water bodies by July 2014 (Fig. 2B). In 2011, significantly higher discharges were noted in OLS3 than in OLS1&2 ($K-W=14.78^*$), whereas in 2014, water flows were similar in all floodplain lakes.

Physicochemical properties of bottom sediments

In this study, samples were collected from relatively new sediment layers that had formed after the restoration. Variations in the physical parameters of the analyzed sediments

	Parameter	Hydrologic stag	e of oxbow lake						Słupia River (RS	3)	
$ \begin{array}{l l l l l l l l l l l l l l l l l l l $		Plesiopotamic			Eupotamic		Parapotamic				
abcabcabiiijpH7.13 \pm 0.067.35 \pm 0.147.56 \pm 0.247.23 \pm 0.437.43 \pm 0.277.42 \pm 0.297.43 \pm 0.237.54 \pm 0.147.65 \pm 0.197.65 \pm 0.29EC450 9 \pm 27.5362.2 \pm 198449.5 \pm 7118325.8 \pm 199.4126.5 \pm 83.9287.4 \pm 133.1178.7 \pm 110.6170 0 \pm 49.8167.6 \pm 54.9158.7 \pm 54.3OM* (%)55.8 \pm 8.735.0 \pm 10.853.3 \pm 8.8°30.4 \pm 18.97.0 \pm 7.72°15.3 \pm 13.21.2.4 \pm 10.22.4 \pm 0.21.1 \pm 0.1BD2.6 \pm 1.71.3 \pm 1.32.3 \pm 0.50.8 \pm 0.42.2 \pm 0.8°1.0 \pm 7.65 \pm 9.41.1 \pm 0.1BD2.6 \pm 1.71.3 \pm 1.32.3 \pm 0.50.8 \pm 0.42.2 \pm 0.8°1.0 \pm 7.65 \pm 9.41.1 \pm 0.1BD2.6 \pm 1.71.3 \pm 1.92.3 \pm 0.50.8 \pm 0.42.2 \pm 0.8°1.0 \pm 7.65 \pm 9.41.0 \pm 7.65 \pm 0.190.27 \pm 0.3TN*18.47 \pm 10.41.2.13 \pm 4.104.2.19 \pm 30.309.19 \pm 3.42°3.43 \pm 2.2854.35 \pm 3.115.70 \pm 3.971.97 \pm 0.58°1.002 \pm 0.13°SRP1.87 \pm 1.041.87 \pm 1.041.87 \pm 1.061.80 \pm 1.561.80 \pm 1.561.01 \pm 0.272.7 \pm 0.3SRP1.87 \pm 1.041.87 \pm 1.061.80 \pm 1.561.80 \pm 1.561.92 \pm 0.13°2.17 \pm 1.061.77 \pm 0.38TOC287 \pm 1.041.87 \pm 1.061.80 \pm 1.561.80 \pm 1.561.01 \pm 0.240.55 \pm 0.160.77 \pm 0.08K1.07 \pm 0.325.1 \pm 1.970.94 \pm 0.44		$\begin{array}{c} \text{OLS1} \\ \text{N} = 24 \end{array}$	$\begin{array}{c} \text{OLS2} \\ \text{N} = 24 \end{array}$	OLS3 $N = 24$	OLS1&2 N=48	OLS 3 $N = 24$	OLS1&2 N=48	$\begin{array}{c} \text{OLS 3} \\ \text{N} = 24 \end{array}$	2007 N=4	2011 N=4	2014 N=4
pH 7.13 ± 0.06 7.35 ± 0.14 7.56 ± 0.24 7.23 ± 0.43 7.43 ± 0.27 7.43 ± 0.23 7.54 ± 0.14 7.63 ± 0.19 7.63 ± 0.29 EC 4509 ± 27.5 362.2 ± 19.8 449.5 ± 71.8 325.8 ± 199.4 126.5 ± 83.9 287.4 ± 133.1 178.7 ± 110.6 1700 ± 49.8 167.6 ± 54.9 158.7 ± 54.3 OM* (%) 55.8 ± 8.7 35.0 ± 10.8 $53.3\pm8.8^\circ$ 30.4 ± 18.9 $7.0\pm7.2^\circ$ 15.3 ± 13.2 12.4 ± 10.2 2.4 ± 0.2 1.2 ± 0.1 1.1 ± 0.1 BD 2.6 ± 1.7 1.3 ± 1.3 2.3 ± 0.5 0.8 ± 0.4 $2.2\pm0.8^\circ$ $1.7+0.8$ 1.5 ± 0.9 4.6 ± 0.4 2.8 ± 0.7 2.7 ± 0.3 TN* 18.47 ± 1.97^h 12.13 ± 4.10 42.19 ± 30.30 9.19 ± 3.42^i 3.43 ± 2.85 4.35 ± 3.11 5.70 ± 3.97 $10.97\pm0.58^\circ$ 10.27 ± 0.08 TN* 18.47 ± 1.97^h 12.13 ± 4.10 42.19 ± 30.30 9.19 ± 3.42^i 3.43 ± 2.85 4.35 ± 3.11 5.70 ± 3.97 $197\pm0.58^\circ$ 10.27 ± 0.013 SRP 1.87 ± 1.04 1.46 ± 0.53 2.56 ± 0.91 0.94 ± 0.43 1.83 ± 1.70 10.0 ± 0.66 1.80 ± 1.35 $10.7\pm0.58^\circ$ $10.7\pm0.13^\circ$ 2.77 ± 0.13 SRP 1.87 ± 1.04 1.46 ± 0.53 $3.50\pm1.126.9$ $1.52\pm4\pm771$ $78.6\pm6.54.4$ 9.8 ± 77.11 $86.1\pm6.8.7$ 2.75 ± 11.5 18.1 ± 8.9 20.6 ± 10.9 SRP 1.87 ± 1.04 1.66 ± 0.53 3.54 ± 4.71 $78.6\pm6.64.6$ 3.35 ± 2.46 3.74 ± 2.32 1.79 ± 0.87 2.26 ± 10.6 1.77 ± 0.84 Na 0.38 ± 0.10 0.55 ± 0.10 0.94 ± 0.26 0.46 ± 0.27 $0.46\pm$		a	p	С	q	в	f	8	<i>h</i>	į	j
EC 450 ± 27.5 362.2 ± 19.8 449.5 ± 71.8 325.8 ± 199.4 126.5 ± 83.9 287.4 ± 133.1 178.7 ± 110.6 170.0 ± 49.8 167.6 ± 54.9 158.7 ± 54.3 OM* (%) 55.8 ± 8.7 35.0 ± 10.8 $53.3 \pm 8.8^{\circ}$ 30.4 ± 18.9 $7.0 \pm 7.2^{\circ}$ 15.3 ± 13.2 12.4 ± 10.2 2.4 ± 0.2 1.1 ± 0.1 DM* (%) 55.8 ± 8.7 35.0 ± 10.8 $53.3 \pm 8.8^{\circ}$ 30.4 ± 18.9 $7.0 \pm 7.2^{\circ}$ 15.3 ± 13.2 12.4 ± 10.2 2.4 ± 0.2 1.2 ± 0.1 1.1 ± 0.1 DD 2.6 ± 1.7 1.3 ± 1.3 2.3 ± 0.5 0.8 ± 0.4 $2.2 \pm 0.8^{\circ}$ 1.7 ± 0.8 1.57 ± 0.9 4.6 ± 0.4 2.8 ± 0.7 2.77 ± 0.3 TN* 18.47 ± 1.97^{h} 12.13 ± 4.10 42.19 ± 30.30 9.19 ± 3.42^{i} 3.43 ± 2.85 4.35 ± 3.11 5.70 ± 3.97 $1.97 \pm 0.58^{\circ}$ $1.02 \pm 0.13^{\circ}$ 2.77 ± 0.3 SRP 1.87 ± 1.04 1.46 ± 0.53 2.56 ± 0.91 0.94 ± 0.43 1.83 ± 1.70 1.00 ± 0.66 1.80 ± 1.35 1.01 ± 0.24 2.8 ± 0.7 2.77 ± 0.3 SRP 1.87 ± 1.04 1.46 ± 0.53 2.55 ± 10.6 0.94 ± 0.43 1.83 ± 1.70 1.00 ± 0.66 1.80 ± 1.35 1.07 ± 0.28 1.07 ± 0.28 1.07 ± 0.28 SRP 1.87 ± 1.04 0.55 ± 0.10 0.94 ± 0.43 1.83 ± 1.70 1.00 ± 0.66 1.80 ± 1.35 $1.01 \pm 0.24 \pm 0.11$ 0.27 ± 0.13 SRP 1.07 ± 0.32 2.05 ± 0.10 0.54 ± 0.20 0.48 ± 0.20 0.48 ± 0.27 0.37 ± 0.14 <td< td=""><td>Hq</td><td>7.13 ± 0.06</td><td>7.35 ± 0.14</td><td>7.56 ± 0.24</td><td>7.23 ± 0.43</td><td>7.43 ± 0.27</td><td>7.42 ± 0.29</td><td>7.43 ± 0.23</td><td>7.54 ± 0.14</td><td>7.63 ± 0.19</td><td>7.63 ± 0.22</td></td<>	Hq	7.13 ± 0.06	7.35 ± 0.14	7.56 ± 0.24	7.23 ± 0.43	7.43 ± 0.27	7.42 ± 0.29	7.43 ± 0.23	7.54 ± 0.14	7.63 ± 0.19	7.63 ± 0.22
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	EC	450.9 ± 27.5	362.2 ± 19.8	449.5 ± 71.8	325.8 ± 199.4	126.5 ± 83.9	287.4 ± 133.1	178.7 ± 110.6	170.0 ± 49.8	167.6 ± 54.9	158.7 ± 54.3
BD 2.6 ± 1.7 1.3 ± 1.3 2.3 ± 0.5 0.8 ± 0.4 2.2 ± 0.8^d 1.7 ± 0.8 1.5 ± 0.9 4.6 ± 0.4 2.8 ± 0.7 2.7 ± 0.3 TN* 18.47 ± 1.97^h $1.2.13\pm4.10$ 42.19 ± 30.30 9.19 ± 3.42^i 3.43 ± 2.85 4.35 ± 3.11 5.70 ± 3.97 1.97 ± 0.58^a 1.02 ± 0.13^d 2.17 ± 1.06 SRP 1.87 ± 1.04 $1.4.6\pm0.53$ 2.56 ± 0.91 0.94 ± 0.43 1.83 ± 1.70 1.00 ± 0.66 1.80 ± 1.35 1.01 ± 0.24 0.52 ± 0.11 0.27 ± 0.08 TOC 287.7 ± 50.0 242.5 ± 110.8 389.1 ± 126.9 $1.52.4\pm47.1$ 78.6 ± 65.4 99.8 ± 73.1 86.1 ± 68.7 27.5 ± 11.5 18.1 ± 8.9 20.6 ± 10.9 TOC 287.7 ± 50.0 242.5 ± 110.8 389.1 ± 126.9 $1.52.4\pm47.1$ 78.6 ± 65.4 99.8 ± 73.1 86.1 ± 68.7 27.5 ± 11.5 18.1 ± 8.9 20.6 ± 10.9 No 0.38 ± 0.10 0.54 ± 0.14 0.5 ± 0.26 0.48 ± 0.27 0.46 ± 0.30 0.37 ± 0.14 0.36 ± 0.17 0.79 ± 0.10 0.76 ± 0.10 0.35 ± 0.06 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.06 ± 1.16 2.72 ± 0.81 2.11 ± 1.06 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.72 ± 0.81 2.72 ± 0.10 0.75 ± 0.10 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.06 ± 1.16 2.72 ± 0.81 2.11 ± 1.06 Mg $1.3.67\pm0.40$ 2.06 ± 0.12 $2.3.9\pm1.460$ 1.826 ± 17.84	0M* (%)	55.8 ± 8.7	35.0 ± 10.8	$53.3\pm8.8^{\circ}$	30.4 ± 18.9	$7.0\pm7.2^{\circ}$	15.3 ± 13.2	12.4 ± 10.2	2.4 ± 0.2	1.2 ± 0.1	1.1 ± 0.1
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	BD	2.6 ± 1.7	1.3 ± 1.3	2.3 ± 0.5	0.8 ± 0.4	2.2 ± 0.8^{d}	1.7 ± 0.8	1.5 ± 0.9	4.6 ± 0.4	2.8 ± 0.7	2.7 ± 0.3
SRP 1.87 ± 1.04 1.46 ± 0.53 2.56 ± 0.91 0.94 ± 0.43 1.83 ± 1.70 1.00 ± 0.66 1.80 ± 1.35 1.01 ± 0.24 0.52 ± 0.11 0.27 ± 0.08 TOC 287.7 ± 50.0 242.5 ± 110.8 389.1 ± 126.9 152.4 ± 47.1 78.6 ± 65.4 99.8 ± 73.1 86.1 ± 68.7 27.5 ± 11.5 18.1 ± 8.9 20.6 ± 10.9 K 1.07 ± 0.32 3.66 ± 0.87 5.1 ± 1.97 2.48 ± 1.06 4.20 ± 2.78 3.35 ± 2.46 3.74 ± 2.32 1.79 ± 0.87 2.20 ± 1.06 1.77 ± 0.84 Na 0.38 ± 0.10 0.54 ± 0.14 0.5 ± 0.26 0.48 ± 0.27 0.46 ± 0.30 0.37 ± 0.14 0.36 ± 0.17 0.79 ± 0.010 0.76 ± 0.10 0.35 ± 0.06 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.60 ± 1.16 2.11 ± 1.00 1.77 ± 0.74 Mg 1.367 ± 0.40 24.61 ± 1.78 15.68 ± 7.87 28.96 ± 19.62 2.33 ± 1.460 18.26 ± 17.84 10.74 ± 3.73 11.22 ± 4.22 5.55 ± 1.30	TN^{*}	$18.47 \pm 1.97^{\rm h}$	12.13 ± 4.10	42.19 ± 30.30	9.19 ± 3.42^{i}	3.43 ± 2.85	4.35 ± 3.11	5.70 ± 3.97	1.97 ± 0.58^{a}	1.02 ± 0.13^d	2.17 ± 1.06
TOC 287.7 ± 50.0 242.5 ± 110.8 389.1 ± 126.9 152.4 ± 47.1 78.6 ± 65.4 99.8 ± 73.1 86.1 ± 68.7 27.5 ± 11.5 18.1 ± 8.9 20.6 ± 10.9 K 1.07 ± 0.32 3.66 ± 0.87 5.1 ± 1.97 2.48 ± 1.06 4.20 ± 2.78 3.35 ± 2.46 3.74 ± 2.32 1.79 ± 0.87 2.20 ± 1.06 1.77 ± 0.84 Na 0.38 ± 0.10 0.54 ± 0.14 0.5 ± 0.26 0.48 ± 0.27 0.46 ± 0.30 0.37 ± 0.14 0.36 ± 0.17 0.79 ± 0.10 0.76 ± 0.10 0.35 ± 0.06 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.60 ± 1.16 2.11 ± 1.00 1.77 ± 0.74 Ca 13.67 ± 0.40 24.61 ± 1.78 15.68 ± 7.87 28.96 ± 19.62 22.37 ± 24.61 23.39 ± 14.60 18.26 ± 17.84 10.74 ± 3.73 11.22 ± 4.22 5.55 ± 1.30	SRP	1.87 ± 1.04	1.46 ± 0.53	2.56 ± 0.91	0.94 ± 0.43	1.83 ± 1.70	1.00 ± 0.66	1.80 ± 1.35	1.01 ± 0.24	0.52 ± 0.11	0.27 ± 0.08
K 1.07 ± 0.32 3.66 ± 0.87 5.1 ± 1.97 2.48 ± 1.06 4.20 ± 2.78 3.35 ± 2.46 3.74 ± 2.32 1.79 ± 0.87 2.20 ± 1.06 1.77 ± 0.84 Na 0.38 ± 0.10 0.54 ± 0.14 0.5 ± 0.26 0.48 ± 0.27 0.46 ± 0.30 0.37 ± 0.14 0.36 ± 0.17 0.79 ± 0.10 0.76 ± 0.10 0.35 ± 0.06 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.60 ± 1.16 2.11 ± 1.00 1.72 ± 0.74 Ca 13.67 ± 0.40 24.61 ± 1.78 15.68 ± 7.87 28.96 ± 19.62 22.37 ± 24.61 23.39 ± 14.60 18.26 ± 17.84 10.74 ± 3.73 11.22 ± 4.22 5.55 ± 1.30	TOC	287.7 ± 50.0	242.5 ± 110.8	389.1 ± 126.9	152.4 ± 47.1	78.6 ± 65.4	99.8 ± 73.1	86.1 ± 68.7	27.5 ± 11.5	18.1 ± 8.9	20.6 ± 10.9
Na 0.38 ± 0.10 0.54 ± 0.14 0.5 ± 0.26 0.48 ± 0.27 0.46 ± 0.30 0.37 ± 0.14 0.36 ± 0.17 0.79 ± 0.10 0.76 ± 0.10 0.35 ± 0.06 Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.60 ± 1.16 2.22 ± 0.81 2.11 ± 1.00 1.72 ± 0.74 Ca 13.67 ± 0.40 24.61 ± 1.78 15.68 ± 7.87 28.96 ± 19.62 22.37 ± 24.61 23.39 ± 14.60 18.26 ± 17.84 10.74 ± 3.73 11.22 ± 4.22 5.55 ± 1.30	K	1.07 ± 0.32	3.66 ± 0.87	5.1 ± 1.97	2.48 ± 1.06	4.20 ± 2.78	3.35 ± 2.46	3.74 ± 2.32	1.79 ± 0.87	2.20 ± 1.06	1.77 ± 0.84
Mg 1.30 ± 0.44 2.06 ± 0.12 3.00 ± 1.71 1.88 ± 0.67 1.99 ± 0.86 2.34 ± 1.23 2.60 ± 1.16 2.11 ± 1.00 1.72 ± 0.74 Ca 13.67 ± 0.40 24.61 ± 1.78 15.68 ± 7.87 28.96 ± 19.62 22.37 ± 24.61 23.39 ± 14.60 18.26 ± 17.84 10.74 ± 3.73 11.22 ± 4.22 5.55 ± 1.30	Na	0.38 ± 0.10	0.54 ± 0.14	0.5 ± 0.26	0.48 ± 0.27	0.46 ± 0.30	0.37 ± 0.14	0.36 ± 0.17	0.79 ± 0.10	0.76 ± 0.10	0.35 ± 0.06
Ca 13.67 ± 0.40 24.61 ± 1.78 15.68 ± 7.87 28.96 ± 19.62 22.37 ± 24.61 23.39 ± 14.60 18.26 ± 17.84 10.74 ± 3.73 11.22 ± 4.22 5.55 ± 1.30	Mg	1.30 ± 0.44	2.06 ± 0.12	3.00 ± 1.71	1.88 ± 0.67	1.99 ± 0.86	2.34 ± 1.23	2.60 ± 1.16	2.22 ± 0.81	2.11 ± 1.00	1.72 ± 0.74
	Ca	13.67 ± 0.40	24.61 ± 1.78	15.68 ± 7.87	28.96 ± 19.62	22.37 ± 24.61	23.39 ± 14.60	18.26 ± 17.84	10.74 ± 3.73	11.22 ± 4.22	5.55 ± 1.30

^{a-e}Statistically significant differences between the studied sites; non parametric Dunn's test, p<0.05

were noted between 2007 and 2014. Statistically significant differences were observed only in bulk density (BD) between OLS2 and the river in 2007 ($K-W=16.17^*$, Table 1).

Plesiopotamal water bodies, i.e., lakes isolated from the river, had the highest particulate organic matter (POM) content. After reconnection with the river, the content of POM decreased visibly when floodplain lakes were flushed with river water (2011). When full connectivity in the RFS was lost again (2014), lentic conditions favored the accumulation of organic debris, which resulted in a doubling of the content of organic matter in lake sediments and relatively high values of TOC (86–152 mg kg⁻¹ DM), TN (5.7–9.2 mg kg⁻¹ DM) and mineralization as expressed by EC (179–326 μ S cm⁻¹).

A nearly twofold increase was observed in the values of TN in the parapotamal phase; however, significant changes in TN in the sediment occurred in OLS1, both during the independent and connection to OLS2 phases. Total nitrogen concentrations were significantly higher in plesiopotamal phase OLS1 than in the river ($K-W=38.67^{**}$) and parapotamal OLS1&2 than in the river in the eupotamal phase ($K-W=13.67^{*}$, Table 1). The higher water residence time contributed to greater sedimentation and deposition of phosphorus.

Taxonomic richness and abundances

A total of 1481 individuals representing 25 taxonomic groups were sampled from the restored oxbow lakes and Słupia River in both years of the different hydrological connectivity phases. Significant differences in the taxonomic composition of invertebrates were noted between the different phases of connection of the RFS. In the plesiopotamal phase, 19 taxa were identified, but in the eupotamal phase, only 5 taxa were identified. Parapotamal-phase benthic fauna were represented by 18 taxa (Table 2). After restoration (eupotamal phase), the number of taxa in the connected lakes OLS1&2 decreased, particularly compared with OLS2 in plesiopotamal water bodies ($K-W=14.00^*$). The greatest increase was observed in OLS3 ($K-W=17.44^{**}$, Table 2).

The density of benthic macroinvertebrates increased significantly ($K-W=15.31^{**}$) between the plesiopotamal phase and after restoration of the eupotamal and parapotamal phases as a result of the project. Invertebrate density increased 27-fold in OLS1&2 and 26-fold in OLS3, and the noted increase was statistically significant (Dunn's test*) in both cases. In the plesiopotamal phase, invertebrate abundance was very high, but 3 years after restoration, invertebrate abundance was low and stable in both OLS1&2 and OLS3. There were no statistically significant differences in the values of the index J', D_{β} and D_{γ} between the studied phases. However, J' and D_{β} were highest in the eupotamal phase (Table 2).

Significant differences in D α values (*K*–*W*=16.28**) were observed between OLS1&2 and the Słupia River in the eupotamal phase during full RFS connectivity (Dunn's test*). In OLS3, D α values increased nearly twofold after restoration. The values of Pielou's evenness index (J') were significantly higher (*K*–*W*=11.25^{*}, Table 3) in the river than in OLS1&2 (Dunn's test*) in the eupotamal phase. Significant differences in the values of J' were not observed between eupotamal oxbow lakes or between OLS3 and the river.

In the plesiopotamal stage of the RFS, the density of invertebrates in the oxbow lakes was determined by similar numbers of Crustacea, Diptera and a slightly smaller number of Oligochaeta (Fig. 3a). A similar pattern was observed in the eupotamal phase but with more than 30-fold lower numbers of whole invertebrate groups. In the parapotamal stage, Oligochaeta were most abundant (Fig. 3b). Changes in the level of connectivity within RFS destructively affected the abundance of Crustacea but had a positive effect on Oligochaeta. In the successive hydrological stages of the RFS, among the trophic guilds, the percentage of shredders

	Plesiopota	mic	Eupotami	ic	Parapotamic		
	Mean	SD	Mean	SD	Mean	SD	
No of taxa*	19	_	5	_	17	_	
Average abundance	2780.2	659.4	80.6	34.0	2367.1	1031.0	
D_{α}^{*}	3.232	0.303	1.815	0.286	2.231	0.690	
J′	0.308	0.251	0.614	0.412	0.160	0.316	
D_{β}	0.171	-	0.200	-	0.172	_	
D_{γ}	0.553	-	0.363	-	0.384	_	

Abundance (indiv. m⁻²), D_{α} =Shannon diversity index, J'=Pielou's evenness index, D_{β} and D_{γ} =Whittaker diversity index

*Statistically significant differences between the studied sites; non parametric Kruskal–Wallis test, p<0.05

Table 2Mean diversity $(\pm SD)$ of invertebrates inhabitingthe oxbow lakes during theconnectivity phases (eachN=72)

$\begin{array}{c c} Plesiopotamic \\ \hline OLS1 & OLS2 \\ N = 24 & N = 24 \\ a & b \end{array}$						Słupia River (R	S)	
$\begin{array}{ccc} OLS1 & OLS2 \\ N=24 & N=24 \\ a & b \end{array}$		Eupotamic		Parapotamic		2007	2011	2014
a b	$\begin{array}{c} \text{OLS } 3\\ \text{N} = 24 \end{array}$	OLS1&2 $N = 48$	OLS 3 N=24	OLS1&2 N=48	OLS 3 N=24	N=4	N =4	N=4
	c	d	в	f	S	<u> </u>	i	j
Richness* 10 13 ^d	16	3 ^b	3f.,g.i	14	15 ^e	10	10 ^e	6
Abundance* $3496.3 \pm 832.1^{d,h}$ 2039.5 ± 32	1^{d} 2804.9±1808.8 ^e	$177.7 \pm 121.4^{a,b,f,g,i}$	$64.2 \pm 86.8^{c,g}$	4889.0 ± 3871.4^{d}	$2133.1 \pm 1722.2^{d,j}$	215.0 ± 100.3^{a}	$160.0\pm60.1^{d,j}$	75.0 ± 32.5^{g}
$D\alpha^{*} \qquad 0.864 \pm 0.156 \qquad 1.205 \pm 0.1$	1.408 ± 0.606	$1.587 \pm 0.224^{e,i}$	0.794 ± 0.347^{i}	1.641 ± 0.565	1.585 ± 0.814	1.458 ± 0.287	$1.923 \pm 0.489^{c,f}$	1.455 ± 0.562
J^* 0.411±0.233 ^h 0.542±0.2	0.606 ± 0.282	0.722 ± 0.323^{i}	0.572 ± 0.501	0.462 ± 0.257	0.499 ± 0.375	0.888 ± 0.198^{a}	$0.835 \pm 0.221^{\circ}$	0.812 ± 0.035
Abundance (indiv. m^{-2}), $D\alpha$ = Shannon dive.	ty index, J' = Pielou's o	evenness index						

by *A. anatim* taxa in the eu (r=0.51-0.7group compri *P. sordens, C P. amnicum,* cies in this gu $p \le 0.05$). The lower abundation were positive mal-phase greater qualitative and remained fair restored floor **Influence of on macroin** In RDA, the axis (F2) exp

ⁱ⁻¹Statistically significant differences between the studied sites; non parametric Dunn's test, p < 0.05

Influence of environmental factors on macroinvertebrate communities

In RDA, the first axis (F1) explained 22.7% and the second axis (F2) explained 19.3% of the variation. In the group of 15 environmental variables tested in RDA, only 5 (flow, TN, SRP, EC and TOC) significantly influenced the quality of the model (Fig. 5a). Preliminary variable selection produced a high correlation between environmental variables and ben-thic macroinvertebrates (r=0.943).

The high values of chemical parameters of sediments were correlated with the plesiopotamal phase and directly proportional to the parapotamal phase. The flow itself was strongly associated with the eupotamal phase. In the RDA analysis, the dispersal of benthic fauna was small, with the exception of Oligochaeta and *A. aquaticus*. The snail *V. contectus* and Heteroptera *Sigara* sp. were characteristic of the parapotamal period; no such characteristic species were identified in other phases of hydrological connectivity (Fig. 5a).

RDA analysis was also performed for the five functional feeding group (Fig. 5b). The first axis (F1) explained 38.4% and the second axis (F2) explained 21.9% of the variation. The same environmental variables were chosen as optimal for the maximum accuracy of the model. High values of SRP

decreased and was substituted by gathering collectors (GC). Predators (Pd) were the least responsive guild with respect to changing hydrological conditions (Fig. 3c). In the absence of plesiopotamal-phase or partial parapotamal-phase connectivity between the oxbows and the river, GC and filtering collectors (FC) constituted a significant percentage of the fauna.

The cluster analysis revealed two significantly different (p < 0.05) groups, referred to as parapotamal- and plesiopotamal-phase groups (Fig. 4). The plesiopotamal water bodies group was represented mostly by Oligochaeta, E. octoculata and, to a lesser extent, V. contectus, A. aquaticus and larvae of the genus Chironomus in floodplain lakes and by A. anatina, P. amnicum and E. danica in the river. All taxa in the eupotamal-phase group were positively correlated (r=0.51-0.77, p < 0.02). The parapotamal floodplain lakes group comprised mainly S. fuliginosa, Lestes sp., N. cinerea, P. sordens, C. dipterum and, to a lesser extent, Heteroptera, P. amnicum, Chironomus sp. and Chaoborus sp. All species in this group were positively correlated (r = 0.46 - 0.56, $p \le 0.05$). The remaining species were characterized by lower abundances. The eupotamal-phase group invertebrates were positively correlated with OLS3, whereas the parapotamal-phase group taxa did not exhibit such preferences. The qualitative and quantitative structures of the benthic fauna remained fairly stable despite considerable changes in the restored floodplain lakes (Fig. 4).

were conducive to higher abundances of GS, whereas an increase in the concentration of TN determined the density of organisms with various modes of obtaining food. With increased concentrations of TOC, the numbers of shredders (Sd) increased.

The generalized linear model (GLM) showed that the increases in the concentrations of TN, SRP and TOC determined the high values of α -diversity (H' index). Simultaneously, the increase in the flow of water through the RFS limited the diversity of benthic fauna in its various parts.

Discussion

Effects of restoration on surface-sediment connectivity

The restoration of hydrological connectivity in the Słupia RFS affected the biotope and parameters of bottom sediments. Those changes were most visible immediately after unblocking of the arm linking the river channel with the floodplain lake. The oxbows were flushed with river water, and fine-grained organic and mineral sediments that had been deposited during the period of isolation from the river were removed.

Significant changes in sediment properties were observed upon the restoration of connectivity (eupotamal phase), and these changes were probably related to the increase in the water content of sediments immediately after reconnection and the increase in the content of POM, TOC and TN (Table 1). The relatively low values of the TOC:TN ratio (<15) in the bottom sediments of oxbow lakes immediately after restoration indicate that phytoplankton were the main source of organic matter (Meyers 2003). The tendency of upstream arms to undergo sedimentation (parapotamal phase) suggests that floodplain lakes naturally evolve into parapotamal habitats when water exchange is inhibited. Those processes contribute to the creation of diverse habitats that accommodate various groups (guilds) of aquatic organisms and enhance biodiversity (Gallardo et al. 2014; Sartori et al. 2015). Changes in the spatial structure and composition of bottom sediments lead to the creation of a wide range of habitats that support the development of benthic fauna. Stagnant water contributes to the sedimentation of fine-grained mineral and organic matter as well as phosphorus deposition. The value of the TOC:TN ratio increases with distance from the river because the accumulation of organic matter in bottom sediments is directly related to biomass production. Floodplain lakes are characterized by

highly variable oxygen levels and qualitative parameters, such as lower nitrate concentrations (due to biosorption and denitrification) and higher ammonia and nitrite levels (due to ammonification and inhibition of nitrification) (Lefebvre et al. 2004).

Although the bottom sediments appear to be uniform, they create significant horizontal and vertical heterogeneities in the physical, chemical, and biological processes in substrata that provide a physical template for distinct niches (Hutchinson 1993). These sedimentary processes include changes in direction and rates of flows, differential deposition of sediment grain sizes and dead organisms, growth and death of roots, burrowing and sediment reworking, and fecal production by benthic consumers. Microhabitats are also created by chemical gradients and microzonation in concentrations of dissolved oxygen, hydrogen sulfide, ammonia, phosphorus, and other critical chemicals (Groffman and Bohlen 1999).

The decline in dissolved oxygen concentrations at the bottom can promote the formation of reduced compounds, such as hydrogen sulphide, resulting in higher adverse (toxic) effects on aquatic animals (Camargo and Alonso 2006). Additionally, the occurrence of toxic algae can significantly contribute to the extensive kills of aquatic animals. Cyanobacteria, dinoflagellates and diatoms appear to be major groups that may be stimulated by inorganic nitrogen pollution. Among the different inorganic nitrogenous compounds $(NH_4^+, NH_3, NO_2^-, NO_3^-)$ that aquatic animals can take up directly from the ambient water, unionized ammonia is the most toxic, while ammonium and nitrate ions are the least toxic.

Water flow and retention time are the key factors responsible for changes in the ecosystems of restored floodplain lakes, as also demonstrated by Tockner et al. (1999). The results of our study show that restoration of full connectivity in the analyzed RFS significantly improved aeration of water, while the decreased retention time of water modified the properties of bottom sediments. The newly created habitat, which was actively flushed with river water, was not favorable for benthic macroinvertebrates. Rapid depletion of particulate organic matter with direct changes in most of the investigated parameters of bottom sediments, including conductivity (EC), total organic carbon (TOC), soluble reactive phosphorus (SRP) and total nitrogen (TN), counteracted the terrestrialization trend of the extant water body. The diversity of macrozoobenthos decreased from 19 taxa in the unaltered water bodies (plesiopotamal phase) to 5 taxa in the reconnected water bodies (eupotamal phase).

Viability of the restoration work

The popularity of lake restoration projects is determined by their long-term effectiveness (Gore et al. 2001; Clarke et al. 2003; Sarriquet et al. 2007). The observed decrease in the nitrogen content of lake sediments may be a transitional process caused by intensified nitrification under aerobic conditions (Chapelle 1993). A periodic decrease in the organic matter content of sediments was also noted. It is difficult to predict whether lake restoration projects will induce permanent or long-lasting changes in the geochemical profile. The changes observed in the third year after the restoration of the eupotamal phase in the analyzed RFS resulted from sediment flushing, whereas in parapotamal habitats, water flow was significantly limited. Total nitrogen concentrations in river sediments increased when water exchange was partially inhibited (Table 1). The above indicates that the restoration of hydrological connectivity in the RFS contributes to the deposition and elimination of nitrogen from floodplain areas (Zalewski 2006). During inundation events in the Słupia River, water circulation in floodplain lakes is intensified, which lowers SRP concentrations due to sediment resuspension. The longer water residence time increases the sedimentation and deposition of phosphorus. Consequently, conservation measures in floodplains should involve the restoration of hydrological connectivity in the RFS and effective management of the entire catchment area.

Effects of restoration on invertebrate assemblages

Habitat conditions and their heterogeneity significantly influence invertebrate populations (Erman and Erman 1984; Merz and Ochikubo Chan 2005). River habitats are modified by changes in the abundance of aquatic macrophytes (Humphries 1996) and/or unobstructed flow of the river channel (Hall et al. 2001). Recent experiments conducted in restored floodplains have demonstrated that unconstrained movement of water from the main stream contributes to habitat heterogeneity and modifies the structure of hydrobionts (Gallardo et al. 2012; Miranda et al. 2014; Paillex et al. 2015). In the present study, a significant increase in the density and diversity of benthic macroinvertebrates was noted in restored oxbow lakes in the Słupia River floodplain over the analyzed period (Table 2). Our findings validate the observations of Ward and Stanford (1995) that floodplains provide supportive habitats for benthic macroinvertebrates. Despite these findings, the restoration of oxbow lakes in the Słupia River floodplain did not lead to a significant increase in the biological diversity of benthic fauna (Table 2). However, the restored hydrological connectivity between the floodplain lakes and river influenced the fluctuation of diversity. The restored floodplain lakes (eupotamal phase) were subjected to two contradictory influences. The inflow of water from the main stream improved water quality in the lakes with respect to oxygen concentration (Obolewski et al. 2016) but despite this it led to a decrease in diversity and abundance of benthic macroinvertebrates due to bottom sediment rinsing. According to Culp et al. (1983), the availability of organic matter is a more critical factor in explaining the composition and density of invertebrate communities than the physical properties of bottom sediments. Therefore, despite an improvement in the physicochemical parameters of the habitat, benthic fauna was influenced by the gradual decrease in organic matter content. The period during which unconstrained water flow was maintained in the oxbow lakes also influenced the abundance and diversity of benthic fauna because the hydrological status of the Słupia River floodplain contributed to a decrease in the diversity and abundance of invertebrates in the river (Table 3).

Changes in the physicochemical properties of sediments in the different phases of connectivity resulted in an increase in the abundance of gathering collectors—GC (Fig. 3c). During the parapotamal and plesiopotamal phases, the percentage of (GC) and filtering collectors (FC) increased. Predators (Pd) did not appear to respond to the hydrological conditions; their presence was associated with the abundance of prey, mainly of the FC guild (Fig. 5b).

Our study was conducted relatively soon after the lake restoration project: therefore, the observed changes in macroinvertebrate communities could be characteristic of the post-colonization phase (Ruhí et al. 2013). The noted processes can, however, be used to identify the main change trends and predict the success of the restoration project, whose main goal was to create stable habitats in the Słupia River floodplain. This is an important consideration because the initially eupotamal habitats were transformed into parapotamal environments, and hydrological connectivity was the main factor influencing the results of this study (Fig. 5a-c). Restricted flow inhibited hydrobiont migration. The observed increase in the abundance of benthic fauna resulted mainly from the increase in the abundance of Oligochaeta, which readily colonizes habitats with high organic matter content and effectively replace less tolerant invertebrate species (Schenková and Helešic 2006).

Changes in outflow and the shorter water retention time in restored habitats also influence the diversity of benthic fauna (Tockner et al. 1999). A decrease in the flow rate modifies habitat conditions in ecosystems and leads to changes in biogeochemical processes (Zalewski 2006) and aquatic fauna (Gallardo et al. 2012; Obolewski et al. 2016). Inhibited flow Fig. 3 Composition of Α invertebrates in relation to the parapotamic hydrological connectivity phase: density of invertebrates taxa (**a** indiv. m^{-2}), relative density eupotamic of invertebrates taxa (b %) and relative density of functional fedding (c %). Sd shredders, GS plesiopotamic grazer scrapers, GC gathering collectors, FC filtering collectors, Pd predators 0 500 1000 1500 2000 2500 3000 indiv. m⁻² Oligochaeta Crustacea Hirudinea Megaloptera Odonata Ephemeroptera Diptera Heteroptera Gastropoda Bivalvia в parapotamic eupotamic plesiopotamic ō 40 100% 20 60 80 Oligochaeta Odonata Crustacea Hirudinea Megaloptera Heteroptera Gastropoda Ephemeroptera Diptera Bivalvia С parapotamic eupotamic plesiopotamic ō 100% 20 40 60 80 Sd GC GS Pd FC

generally decreases the density and abundance of invertebrate species (Meyer and Meyer 2000; Boulton 2003) and leads to changes in the size and age structure of hydrobiont populations (Lake 2003). The results of our study indicate that floodplain ecosystems are more likely to thrive in RFS characterized by a partial parapotamal rather than parapotamal phase. Other authors have observed that the main goal of floodplain restoration projects should be to "refresh" the biotope with river water (Ward et al. 2002; Paillex et al. 2015), consistent with our findings.

Fig. 4 Two-way cluster analysis dendrogram of 14 plots and 25 macroinvertebrate taxa. Each square represents relative abundance by column (darker is more abundant). The plot clusters are indicated at the branch tips with vertical black lines and at the node with numbers. The site clusters are indicated with red color. Codes for organism names: OLI, Oligochaeta; Ase aqu, Asellus aquaticus; Gam_ pul, Gammarus pulex; Glos_ com, Glossiphonia complanata; Erp_oct, Erpobdella octoculata; Hel sp., *Hellobdela* sp.; Sia ful, Sialis fuliginosa; Les_sp., Lestes sp.; Clo_dip., Cloëon dipterum; Eph_dan, Ephemera danica; Chir_sp., Chironomus sp.; Chao_sp., Chaoborus sp.; Pol_sor, Polypedilum sordens; Ser_sp., Sergentia sp.; Pro_sp., Procladius sp.; Cor_sp., Corixa sp.; Hes_sp., Hesperocorixa sp.; Nep_cin, Nepa cinera; Cym_ sp., Cymbiodyta sp.; Sig_sp., Sigara sp.; Dyt_mar, Dytiscus marginalis; Viv con, Viviparus contectus; Ano_ana, Anodonta anatina; Pis_sp., Pisidum sp.; Sph_sp., Sphaerium sp.



Conclusions

The results of this study validate the hypothesis that the restoration of hydrological connectivity in RFS influences the properties of bottom sediments in floodplain lakes. The conservation project also exerted a significant influence on benthic fauna, but the abundance of macroinvertebrate taxa was more correlated with hydrological connectivity than sediment properties. The gradual loss of hydrological connectivity led to significant changes in the structure and chemical composition of bottom sediments and increased the abundance and diversity of macroinvertebrates. Where possible river valleys should be allowed to flood periodically and thus naturally create one extensive ecological system. Our findings indicate that floodplain lakes should be periodically connected with their rivers to "refresh" their biotope.

Our study reveals that conservation projects aimed at restoring connectivity between floodplain areas and the main

stream are unable to prevent natural erosion and accumulation processes or alter the dynamics of a river's hydrological regime. Restoration projects should be developed in view of long-term natural hydrodynamic processes, which determine the success of conservation measures. This study documents the ecological consequences of a restoration project over a period of only 5 years, but the geochemical profile of the floodplain could undergo significant changes in the long term. Such changes are influenced by the inflow of dissolved substances from the catchment and other physical factors (inundation, erosion of the river bed). Therefore, to ensure maximum sustainability, projects that restore hydrological connectivity in a floodplain should also involve effective management of catchment areas, including limited fertilizer use, creation of buffer zones and protection of littoral vegetation.

Fig. 5 The triplot of redudency analysis (a) for the environmental variables, samples and composition of benthofauna and biplot for the environmental variables and functional feeding groups (b). Codes for organism names the same as for Fig. 4. General linear model (c) for the environmental variables and distribution of D α



Compliance with ethical standards

Conflict of interest The authors have declared no conflict of interest.

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Appendix

See Table 4.

 Table 4
 Relative density (%) of benthofauna inhabiting the studied oxbow lakes and Słupia River before (plesiopotamic phase) and after restoration (eupotamic and parapotamic phases)

	FFG	Connectivity phase							2007	2011	2014
		Plesiop	otamic		Eupotamic		Parapotamic				
		OLS1	OLS2	OLS3	OLS1&2	OLS3	OLS1&2	OLS3	River S	Słupia	
Oligochaeta	GC	3.0	23.1	35.7	55.7	76.9	66.3	79.6	4.7	3.2	6.7
Asellus aquaticus	Sd	28.4	35.8	42.9	30.4	0.0	0.1	1.4	4.7	3.2	0.0
Gammarus pulex	Sd	0.0	0.0	0.5	0.0	0.0	0.0	0.0	0.0	3.2	0.0
Glossiphonia complanata	Pd	0.4	0.0	2.3	0.0	0.0	0.1	3.2	0.0	0.0	0.0
Erpobdella octoculata	Pd	0.8	1.2	1.0	0.0	15.4	1.0	2.8	2.3	0.0	0.0
Hellobdela sp.	Pd	0.0	0.2	2.3	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Sialis fuliginosa	Pd	0.6	0.5	2.1	0.0	0.0	0.5	1.9	0.0	0.0	0.0
Libellula quadrimaculata	Pd	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.3	6.5	6.7
Lestes sp.	Pd	0.0	0.0	0.0	0.0	0.0	0.0	1.4	0.0	3.2	0.0
Cloëon dipterum	GS	4.0	5.5	0.0	0.0	0.0	4.1	1.4	0.0	22.6	6.7
Ephemera danica	GS	0.0	0.0	0.3	0.0	0.0	0.0	0.0	16.3	0.0	0.0
Chironomus sp.	GC	55.5	19.0	2.4	13.9	0.0	23.2	3.2	25.6	16.1	33.3
Chaoborus sp.	Pd	0.0	0.5	0.0	0.0	0.0	0.2	0.0	0.0	0.0	0.0
Polypedilum sordens	GS	6.8	0.0	0.0	0.0	0.0	0.0	0.7	7.0	0.0	0.0
Procladius sp.	Pd	0.0	9.4	2.4	0.0	0.0	0.0	0.0	23.3	0.0	0.0
Sergentia sp.	GC	0.0	1.0	5.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Corixa sp.	Pd	0.3	0.2	0.2	0.0	0.0	0.9	0.2	0.0	0.0	0.0
Hesperocorixa sp.	Pd	0.0	0.0	0.0	0.0	0.0	0.1	0.0	0.0	0.0	0.0
Nepa cinera	Pd	0.0	0.0	0.2	0.0	0.0	0.0	1.6	0.0	0.0	0.0
<i>Sigara</i> sp.	Pd	0.0	0.0	0.0	0.0	0.0	0.4	0.9	0.0	0.0	0.0
Cymbiodyta sp.	Pd	0.0	0.2	0.9	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Dytiscus marginalis	Pd	0.0	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Viviaparus contectus	Sd	0.0	0.0	0.0	0.0	7.7	2.2	0.7	2.3	0.0	0.0
Anodonta anatina	FC	0.0	0.0	0.0	0.0	0.0	0.1	0.0	0.0	9.7	0.0
Pisidium sp.	FC	0.0	0.0	0.0	0.0	0.0	0.7	0.5	0.0	16.1	26.7
Sphaerium sp.	FC	0.3	3.4	1.4	0.0	0.0	0.0	0.5	11.6	16.1	20.0

FFG functional feeding group (Acronyms: Sd shredders, GS grazer scrapers, GC gathering collectors, FC filtering collectors, Pd predators)

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